

# Package ‘IDSpatialStats’

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**Title** Estimate Global Clustering in Infectious Disease

**License** GPL ( $\geq 2$ )

**Description** Implements various novel and standard clustering statistics and other analyses useful for understanding the spread of infectious disease.

**RoxygenNote** 7.3.1

**Encoding** UTF-8

**LazyData** true

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**NeedsCompilation** yes

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DengueSimR01	<i>Simulated dataset of dengue transmission with basic reproductive number of 1</i>
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---

## Description

Dataset simulated using an agent based model with a spatially heterogeneous population structure. Infectious agents were introduced resulting in agent to agent transmission. The distance between successive cases in a transmission chain were randomly drawn from a uniform distribution  $U(0,100)$ . Each infectious agent resulted in a single transmission to another agent after a delay of 15 days, reflecting the generation time of dengue. There are 11 transmission chains, each with a different genotype. The genotypes are subdivided into four serotypes.

**Usage**

DengueSimR01

**Format**

Matrix with five columns representing the X and Y coordinates of infected individuals, the time of infection, the genotype of the infecting pathogen and the serotype of the infecting pathogen.

**Author(s)**

Justin Lessler and Henrik Salje

---

DengueSimR02	<i>Simulated dataset of dengue cases with basic reproductive number of 2</i>
--------------	--

---

**Description**

Dataset simulated using an agent based model with a spatially heterogeneous population structure. Infectious agents were introduced resulting in agent to agent transmission. The distance between successive cases in a transmission chain were randomly drawn from a uniform distribution  $U(0,100)$ . Each infectious agent resulted in transmissions to two other agents after a delay of 15 days, reflecting the generation time of dengue. There are 11 transmission chains, each with a different genotype. The genotypes are subdivided into four serotypes.

**Usage**

DengueSimR02

**Format**

Matrix with five columns representing the X and Y coordinates of infected individuals, the time of infection, the genotype of the infecting pathogen and the serotype of the infecting pathogen.

**Author(s)**

Justin Lessler and Henrik Salje

DengueSimRepresentative

*Simulated dataset of dengue cases with representative underlying population*

---

### **Description**

Dataset simulated using an agent based model with a spatially heterogeneous population structure. Infectious agents were introduced resulting in agent to agent transmission. The distance between successive cases in a transmission chain were randomly drawn from a uniform distribution  $U(0,100)$ . Each infectious agent resulted in transmissions to two other agents after a delay of 15 days, reflecting the generation time of dengue. There are 11 transmission chains, each with a different genotype. The genotypes are subdivided into four serotypes. 500 randomly selected individuals from the underlying population also included.

### **Usage**

DengueSimRepresentative

### **Format**

Matrix with five columns representing the X and Y coordinates of infected individuals, the time of infection, the genotype of the infecting pathogen and the serotype of the infecting pathogen. Individuals representative from the underlying population have '-999' for time, genotype and serotype.

### **Author(s)**

Justin Lessler and Henrik Salje

---

est.transdist

*Estimate transmission distance*

---

### **Description**

this function estimates the mean transmission distance of an epidemic when given the locations and times of symptomatic cases and the mean and standard deviation of the generation time of the infecting pathogen

### **Usage**

```
est.transdist(  
  epi.data,  
  gen.t.mean,  
  gen.t.sd,  
  t1,  
  max.sep,
```

```

    max.dist,
    n.transtree.reps = 100,
    theta.weights = NULL
  )

```

### Arguments

<code>epi.data</code>	a three-column matrix giving the coordinates (x and y) and time of infection (t) for all cases in an epidemic (columns must be in x, y, t order)
<code>gen.t.mean</code>	mean generation time of the infecting pathogen
<code>gen.t.sd</code>	standard deviation of generation time of the infecting pathogen
<code>t1</code>	time step to begin estimation of transmission distance
<code>max.sep</code>	maximum number of time steps allowed between two cases (passed to the <code>get.transdist.theta</code> function)
<code>max.dist</code>	maximum spatial distance between two cases considered in calculation
<code>n.transtree.reps</code>	number of time to simulate transmission trees when estimating the weights of theta (passed to the <code>est.transdist.theta.weights</code> function, default = 10). Warning: higher values of this parameter cause significant increases in computation time.
<code>theta.weights</code>	use external matrix of theta weights. If NULL (default) the matrix of theta weights is automatically estimated by calling the <code>est.transdist.theta.weights</code> function

### Value

a list containing the estimated mean distance of the transmission kernel (`mu.est`) and its standard deviation (`sigma.est`). Bounded estimates (`bound.mu.est` and `bound.sigma.est`) are also given for when the assumption of equal mean and standard deviation is violated.

### Author(s)

John Giles, Justin Lessler, and Henrik Salje

### References

Salje H, Cummings DAT and Lessler J (2016). “Estimating infectious disease transmission distances using the overall distribution of cases.” *Epidemics*, 17, pp. 10–18. ISSN 1755-4365, doi: [10.1016/j.epidem.2016.10.001](https://doi.org/10.1016/j.epidem.2016.10.001).

### See Also

Other est.wt: [est.wt.matrix\(\)](#), [est.wt.matrix.weights\(\)](#)

Other transdist: [est.transdist.bootstrap.ci\(\)](#), [est.transdist.temporal\(\)](#), [est.transdist.temporal.bootstrap\(\)](#), [est.transdist.theta.weights\(\)](#), [get.transdist.theta\(\)](#)

**Examples**

```

set.seed(123)

# Exponentially distributed transmission kernel with mean and standard deviation = 100
dist.func <- alist(n=1, a=1/100, rexp(n, a))

# Simulate epidemic
a <- sim.epidemic(R=1.5,
                 gen.t.mean=7,
                 gen.t.sd=2,
                 min.cases=50,
                 tot.generations=12,
                 trans.kern.func=dist.func)

# Estimate mean and standard deviation of transmission kernel
b <- est.transdist(eps.data=a,
                 gen.t.mean=7,
                 gen.t.sd=2,
                 t1=0,
                 max.sep=1e10,
                 max.dist=1e10,
                 n.transtree.reps=10)
b

```

---

```
est.transdist.bootstrap.ci
```

*Bootstrap mean transmission distance values*

---

**Description**

Runs `est.transdist` on multiple bootstraps of the data and calculates confidence intervals for the mean transmission distance.

**Usage**

```

est.transdist.bootstrap.ci(
  eps.data,
  gen.t.mean,
  gen.t.sd,
  t1,
  max.sep,
  max.dist,
  n.transtree.reps = 100,
  mean.equals.sd = FALSE,
  theta.weights = NULL,

```

```

boot.iter,
ci.low = 0.025,
ci.high = 0.975,
parallel = FALSE,
n.cores = NULL
)

```

### Arguments

<code>epi.data</code>	a three-column matrix giving the coordinates (x and y) and time of infection (t) for all cases in an epidemic (columns must be in x, y, t order)
<code>gen.t.mean</code>	mean generation time of the infecting pathogen
<code>gen.t.sd</code>	standard deviation of generation time of the infecting pathogen
<code>t1</code>	time step to begin estimation of transmission distance
<code>max.sep</code>	maximum number of time steps allowed between two cases (passed to the <code>get.transdist.theta</code> function)
<code>max.dist</code>	maximum spatial distance between two cases considered in calculation
<code>n.transtree.reps</code>	number of time to simulate transmission trees when estimating the weights of theta (passed to the <code>est.transdist.theta.weights</code> function, default = 10). Warning: higher values of this parameter cause significant increases in computation time.
<code>mean.equals.sd</code>	logical term indicating if the mean and standard deviation of the transmission kernel are expected to be equal (default = FALSE)
<code>theta.weights</code>	use external matrix of theta weights. If NULL (default) the matrix of theta weights is automatically estimated by calling the <code>est.transdist.theta.weights</code> function
<code>boot.iter</code>	the number of bootstrapped iterations to perform
<code>ci.low</code>	low end of the confidence interval (default = 0.025)
<code>ci.high</code>	high end of the confidence interval (default = 0.975)
<code>parallel</code>	run bootstraps in parallel (default = FALSE)
<code>n.cores</code>	number of cores to use when <code>parallel = TRUE</code> (default = NULL, which uses half the available cores)

### Value

a list object containing the point estimate for mean transmission distance and low and high bootstrapped confidence intervals

### Author(s)

John Giles, Justin Lessler, and Henrik Salje

**References**

Salje H, Cummings DAT and Lessler J (2016). “Estimating infectious disease transmission distances using the overall distribution of cases.” *Epidemics*, 17, pp. 10–18. ISSN 1755-4365, doi: [10.1016/j.epidem.2016.10.001](https://doi.org/10.1016/j.epidem.2016.10.001).

**See Also**

Other transdist: [est.transdist\(\)](#), [est.transdist.temporal\(\)](#), [est.transdist.temporal.bootstrap.ci\(\)](#), [est.transdist.theta.weights\(\)](#), [get.transdist.theta\(\)](#)

**Examples**

```
set.seed(1)

# Exponentially distributed transmission kernel with mean and standard deviation = 100
dist.func <- alist(n=1, a=1/100, rexp(n, a))

# Simulate epidemic
a <- sim.epidemic(R=2.5,
                 gen.t.mean=7,
                 gen.t.sd=2,
                 min.cases=20,
                 tot.generations=5,
                 trans.kern.func=dist.func)

# Estimate mean transmission kernel and its bootstrapped confidence intervals
b <- est.transdist.bootstrap.ci(epi.data=a,
                              gen.t.mean=7,
                              gen.t.sd=2,
                              t1=0,
                              max.sep=1e10,
                              max.dist=1e10,
                              n.transtree.reps=10,
                              mean.equals.sd=TRUE,
                              boot.iter=10,
                              ci.low=0.025,
                              ci.high=0.975,
                              n.cores=2)

b
```

---

`est.transdist.temporal`

*Change in mean transmission distance over time*

---



**Description**

Estimates the change in mean transmission distance over the duration of the epidemic by running `est.transdist` on all cases occurring up to each time point.

**Usage**

```
est.transdist.temporal(  
  epi.data,  
  gen.t.mean,  
  gen.t.sd,  
  t1,  
  max.sep,  
  max.dist,  
  n.transtree.reps = 10,  
  mean.equals.sd = FALSE,  
  theta.weights = NULL,  
  parallel = FALSE,  
  n.cores = NULL  
)
```

**Arguments**

<code>epi.data</code>	a three-column matrix giving the coordinates (x and y) and time of infection (t for all cases in an epidemic (columns must be in x, y, t order)
<code>gen.t.mean</code>	mean generation time of the infecting pathogen
<code>gen.t.sd</code>	standard deviation of generation time of the infecting pathogen
<code>t1</code>	time step to begin estimation of transmission distance
<code>max.sep</code>	maximum number of time steps allowed between two cases (passed to the <code>est.transdist.theta</code> function)
<code>max.dist</code>	maximum spatial distance between two cases considered in calculation
<code>n.transtree.reps</code>	number of time to simulate transmission trees when estimating the weights of theta (passed to the <code>est.transdist.theta.weights</code> function, default = 10). Higher values of this parameter cause significant increases in computation time.
<code>mean.equals.sd</code>	logical term indicating if the mean and standard deviation of the transmission kernel are expected to be equal (default = FALSE)
<code>theta.weights</code>	use external matrix of theta weights. If NULL (default) the matrix of theta weights is automatically estimated by calling the <code>est.transdist.theta.weights</code> function
<code>parallel</code>	run time steps in parallel (default = FALSE)
<code>n.cores</code>	number of cores to use when <code>parallel = TRUE</code> (default = NULL, which uses half the available cores)

**Value**

a numeric matrix containing the point estimate for mean transmission distance for each unique time step of the epidemic and the sample size  $n$  used to make the estimate NAs are returned for time steps which contain fewer than three cases

**Author(s)**

John Giles, Justin Lessler, and Henrik Salje

**References**

Salje H, Cummings DAT and Lessler J (2016). “Estimating infectious disease transmission distances using the overall distribution of cases.” *Epidemics*, 17, pp. 10–18. ISSN 1755-4365, doi: [10.1016/j.epidem.2016.10.001](https://doi.org/10.1016/j.epidem.2016.10.001).

**See Also**

Other transdist: [est.transdist\(\)](#), [est.transdist.bootstrap.ci\(\)](#), [est.transdist.temporal.bootstrap.ci\(\)](#), [est.transdist.theta.weights\(\)](#), [get.transdist.theta\(\)](#)

**Examples**

```
set.seed(123)

# Exponentially distributed transmission kernel with mean and standard deviation = 100
dist.func <- alist(n=1, a=1/100, rexp(n, a))

# Simulate epidemic
a <- sim.epidemic(R=2,
                 gen.t.mean=7,
                 gen.t.sd=2,
                 tot.generations=7,
                 min.cases=30,
                 trans.kern.func=dist.func)

a <- a[sample(1:nrow(a), 50),] # subsample a to 50 observations

# Estimate mean transmission kernel over time
b <- est.transdist.temporal(eps.data=a,
                          gen.t.mean=7,
                          gen.t.sd=2,
                          t1=0,
                          max.sep=1e10,
                          max.dist=1e10,
                          n.transtree.reps=5,
                          mean.equals.sd=TRUE,
                          n.cores=2)

b

plot(b[,2], pch=19, col='grey', ylim=c(min(b[,2], na.rm=TRUE), max(b[,2], na.rm=TRUE)),
```

```

      xlab='Time step', ylab='Estimated mean of transmission kernel')
abline(h=100, col='red', lty=2)
axis(3, b[,2])

low <- loess(b[,2] ~ as.vector(1:length(b[,2])))
low <- predict(low, newdata=data.frame(as.vector(1:length(b[,2])))
lines(low, lwd=3, col='blue')

```

---

```
est.transdist.temporal.bootstrap.ci
```

*Bootstrapped confidence intervals for the change in mean transmission distance over time*

---

### Description

Estimates bootstrapped confidence intervals for the mean transmission distance over the duration of the epidemic by running `est.transdist` on all cases occurring up to each time point.

### Usage

```

est.transdist.temporal.bootstrap.ci(
  epi.data,
  gen.t.mean,
  gen.t.sd,
  t1,
  max.sep,
  max.dist,
  n.transtree.reps = 100,
  mean.equals.sd = FALSE,
  theta.weights = NULL,
  boot.iter,
  ci.low = 0.025,
  ci.high = 0.975,
  parallel = FALSE,
  n.cores = NULL
)

```

### Arguments

<code>epi.data</code>	a three-column matrix giving the coordinates (x and y) and time of infection (t for all cases in an epidemic (columns must be in x, y, t order)
<code>gen.t.mean</code>	mean generation time of the infecting pathogen
<code>gen.t.sd</code>	standard deviation of generation time of the infecting pathogen
<code>t1</code>	time step to begin estimation of transmission distance

<code>max.sep</code>	maximum number of time steps allowed between two cases (passed to the <code>get.transdist.theta</code> function)
<code>max.dist</code>	maximum spatial distance between two cases considered in calculation
<code>n.transtree.reps</code>	number of time to simulate transmission trees when estimating the weights of <code>theta</code> (passed to the <code>est.transdist.theta.weights</code> function, default = 10). Warning: higher values of this parameter cause significant increases in computation time.
<code>mean.equals.sd</code>	logical term indicating if the mean and standard deviation of the transmission kernel are expected to be equal (default = FALSE)
<code>theta.weights</code>	use external matrix of <code>theta</code> weights. If NULL (default) the matrix of <code>theta</code> weights is automatically estimated by calling the <code>est.transdist.theta.weights</code> function
<code>boot.iter</code>	the number of bootstrapped iterations to perform
<code>ci.low</code>	low end of the confidence interval (default = 0.025)
<code>ci.high</code>	high end of the confidence interval (default = 0.975)
<code>parallel</code>	run bootstraps in parallel (default = FALSE)
<code>n.cores</code>	number of cores to use when <code>parallel = TRUE</code> (default = NULL, which uses half the available cores)

**Value**

a four-column numeric matrix containing the point estimate for mean transmission distance, low and high bootstrapped confidence intervals, and the sample size up to each time step

**Author(s)**

John Giles, Justin Lessler, and Henrik Salje

**References**

Salje H, Cummings DAT and Lessler J (2016). “Estimating infectious disease transmission distances using the overall distribution of cases.” *Epidemics*, 17, pp. 10–18. ISSN 1755-4365, doi: [10.1016/j.epidem.2016.10.001](https://doi.org/10.1016/j.epidem.2016.10.001).

**See Also**

Other `transdist`: `est.transdist()`, `est.transdist.bootstrap.ci()`, `est.transdist.temporal()`, `est.transdist.theta.weights()`, `get.transdist.theta()`

**Examples**

```
set.seed(123)

# Exponentially distributed transmission kernel with mean and standard deviation = 100
dist.func <- alist(n=1, a=1/100, rexp(n, a))
```

```

# Simulate epidemic
a <- sim.epidemic(R=2,
                 gen.t.mean=7,
                 gen.t.sd=2,
                 tot.generations=8,
                 min.cases=30,
                 trans.kern.func=dist.func)

a <- a[sample(1:nrow(a), 70),] # subsample a to 70 observations

# Estimate change in mean transmission kernel over time with confidence intervals
b <- est.transdist.temporal.bootstrap.ci(epi.data=a,
                                       gen.t.mean=7,
                                       gen.t.sd=2,
                                       t1=0,
                                       max.sep=1e10,
                                       max.dist=1e10,
                                       n.transtree.reps=10,
                                       mean.equals.sd=TRUE,
                                       boot.iter=10,
                                       ci.low=0.025,
                                       ci.high=0.975,
                                       n.cores=2)

plot(b[,2], pch=19, col='grey', ylim=c(min(b[,1:3], na.rm=TRUE), max(b[,2:4], na.rm=TRUE)),
     xlab='Time step', ylab='Estimated mean of transmission kernel')
abline(h=100, col='red', lty=2)
axis(3, 1:nrow(b), b[,5])

low <- loess(b[,2] ~ as.vector(1:nrow(b)), span=1)
low <- predict(low, newdata=data.frame(as.vector(1:nrow(b))))
lines(low, lwd=3, col='blue')

for(i in 3:4) {
  low <- loess(b[,i] ~ as.vector(1:nrow(b)), span=1)
  low <- predict(low, newdata=data.frame(as.vector(1:nrow(b))))
  lines(low, lty=2, lwd=3, col='blue')
}

```

---

```
est.transdist.theta.weights
```

*Estimate transmission distance theta values by replication*

---

### Description

This function estimates the weight of each theta value by performing a user defined number of replications with the `get.transdist.theta` function. The weights of each theta are calculated as

the number of simulations in which a case at time `t1` and `t2` are separated by `theta` transmission events.

### Usage

```
est.transdist.theta.weights(case.times, gen.t.mean, t.density, t1, n.rep = 100)
```

### Arguments

<code>case.times</code>	a vector giving the occurrence time for each case
<code>gen.t.mean</code>	the mean generation time of the infecting pathogen
<code>t.density</code>	a vector giving the generation time density of the infecting pathogen
<code>t1</code>	time step to begin simulation
<code>n.rep</code>	number of replications in the simulation (default = 100)

### Value

a three-dimensional array containing the mean normalized `theta` weights estimated across all replications

### Author(s)

John Giles, Justin Lessler, and Henrik Salje

### References

Salje H, Cummings DAT and Lessler J (2016). “Estimating infectious disease transmission distances using the overall distribution of cases.” *Epidemics*, 17, pp. 10–18. ISSN 1755-4365, doi: [10.1016/j.epidem.2016.10.001](https://doi.org/10.1016/j.epidem.2016.10.001).

### See Also

Other `transdist`: [est.transdist\(\)](#), [est.transdist.bootstrap.ci\(\)](#), [est.transdist.temporal\(\)](#), [est.transdist.temporal.bootstrap.ci\(\)](#), [get.transdist.theta\(\)](#)

### Examples

```
set.seed(1)

gen.t.mean <- 7
gen.t.sd <- 2
t1 <- 0

# Normally distributed transmission kernel with mean and standard deviation = 100
dist.func <- alist(n=1, a=1/100, rexp(n, a))

# Simulate epidemic
a <- sim.epidemic(R=5,
                  gen.t.mean=gen.t.mean,
```

```

        gen.t.sd=gen.t.sd,
        min.cases=5,
        tot.generations=3,
        trans.kern.func=dist.func)

# Get case times
a <- a[order(a[,3]),]
case.times <- round(a[,3])
unique.times <- unique(case.times)
ntimes <- length(unique.times)

# Generation time distribution
max.t <- round(max(unique.times) - t1) - 1
n.step <- round(max.t/gen.t.mean)
gen <- rep(0, max.t*2)
for (i in 1:n.step){gen <- gen + dnorm(1:(max.t*2), gen.t.mean*i, gen.t.sd*i)}
gen[1] <- 0 # No instantaneous infections
t.density <- gen/sum(gen)

# Estimation of theta weights matrix
b <- est.transdist.theta.weights(case.times=case.times,
                                n.rep=3,
                                gen.t.mean=gen.t.mean,
                                t1=t1,
                                t.density=t.density)

```

---

est.wt.matrix

*Calculate the Infector-Infectee Wallinga-Teunis matrix*


---

## Description

A function which takes the time of each case occurrence, the generation time distribution of the infecting pathogen, and the matrix of basic Wallinga-Teunis weights and estimates the probability that an infectee occurring time step  $j$  (columns) was infected by a case occurring at time  $i$  (rows).

## Usage

```
est.wt.matrix(case.times, gen.t.dist, basic.wt.weights = NULL)
```

## Arguments

`case.times` a vector giving the occurrence time for each case

`gen.t.dist` a vector giving the generation time distribution for the infecting pathogen

`basic.wt.weights` a matrix giving the basic normalized Wallinga-Teunis weights for each time step (output from the `est.wt.matrix.weights` function). If this argument is `NULL` (the default), the basic Wallinga-Teunis matrix will be calculated automatically.

**Value**

a numerical matrix with the number of columns and rows equal to the number of cases in the epidemic

**Author(s)**

John Giles, Justin Lessler, and Henrik Salje

**References**

Salje H, Cummings DAT and Lessler J (2016). “Estimating infectious disease transmission distances using the overall distribution of cases.” *Epidemics*, 17, pp. 10–18. ISSN 1755-4365, doi: [10.1016/j.epidem.2016.10.001](https://doi.org/10.1016/j.epidem.2016.10.001).

**See Also**

Other est.wt: [est.transdist\(\)](#), [est.wt.matrix.weights\(\)](#)

**Examples**

```
case.times <- c(1,2,2,3,3)
gen <- c(0, 2/3, 1/3, 0, 0)
t.density <- gen/sum(gen)

a <- est.wt.matrix(case.times=case.times, gen.t.dist=t.density)
```

---

est.wt.matrix.weights *Estimate matrix of basic Wallinga-Teunis weights*

---

**Description**

A function called by `est.wt.matrix`, which calculates the basic weights in the Wallinga-Teunis matrix given the time of each case occurrence and the generation time distribution of the pathogen. Code adapted from the **R0** package.

**Usage**

```
est.wt.matrix.weights(case.times, gen.t.dist)
```

**Arguments**

<code>case.times</code>	a vector giving the occurrence time for each case
<code>gen.t.dist</code>	a vector giving the generation time distribution for the infecting pathogen



**Value**

a numerical matrix with the number of columns and rows equal to the number of time steps in the epidemic

**Author(s)**

John Giles, Justin Lessler, and Henrik Salje

**References**

Boelle P and Obadia T (2015). R0: Estimation of R0 and Real-Time Reproduction Number from Epidemics. R package version 1.2-6, <https://CRAN.R-project.org/package=R0>.

Salje H, Cummings DAT and Lessler J (2016). “Estimating infectious disease transmission distances using the overall distribution of cases.” *Epidemics*, 17, pp. 10–18. ISSN 1755-4365, doi: [10.1016/j.epidem.2016.10.001](https://doi.org/10.1016/j.epidem.2016.10.001).

**See Also**

Other est.wt: [est.transdist\(\)](#), [est.wt.matrix\(\)](#)

**Examples**

```
case.times <- c(1,2,2,3,3)
gen <- c(0, 2/3, 1/3, 0, 0)
t.density <- gen/sum(gen)

a <- est.wt.matrix.weights(case.times=case.times, gen.t.dist=t.density)
```

---

get.cross.K

*Cross type K function using homotypic and heterotypic case types*

---

**Description**

A wrapper function of the [Kcross](#) function from the **spatstat.explore** package (Baddeley et al. 2016) that takes epidemiological data used by **IDSpatialStats** functions and calculates the cross type K-function based on user defined case type homology

**Usage**

```
get.cross.K(epi.data, type, hom, het = NULL, r = NULL, correction = "none")
```

**Arguments**

epi.data	a three-column numerical matrix that contains coordinates (x and y) for each case and information on case type (e.g. genotype or serotype). First two columns must be x and y
type	an integer giving the column that contains information on case type. Must be an integer or a character
hom	a scalar or vector giving the homotypic case type(s). Equivalent to the 'j' point type used in the cross K function. Must be an integer or character
het	a scalar or vector giving the heterotypic case type(s). Equivalent to the 'i' point type used in the cross K function. The default is NULL, which uses any case type not defined in the hom argument as heterotypic. Must be an integer or a character
r	a numeric vector giving the spatial distances
correction	type of edge correction to be applied (default set to 'none'). See the <a href="#">Kcross</a> function in the <b>spatstat.explore</b> package for more details

**Value**

a data frame with a minimum of three columns giving the radius (r), the theoretical value of the K function for a Poisson process (theo), and value of the K function evaluated at radius r. The column name gives the type of edge correction used

**Author(s)**

John Giles

**References**

Baddeley A, Rubak E, and Turner R. (2016). "Spatial Point Patterns: Methodology and Applications with R". CRC Press.

**Examples**

```
data(DengueSimR01)

k <- get.cross.K(epi.data=DengueSimR01, type=5, hom=2, het=NULL, r=NULL, correction='border')

plot(k[,2], type='l', col='red', lty=2, xlab='r', ylab='cross K function')
lines(k$border)
```

---

get.cross.PCF	<i>Cross type Pair Correlation Function using homotypic and heterotypic case types</i>
---------------	--

---

### Description

A wrapper function of the [pcf](#) function from the **spatstat.explore** package (Baddeley et al. 2016) that takes epidemiological data used by **IDSpatialStats** functions and calculates the cross type Pair Correlation Function based on user defined case type homology

### Usage

```
get.cross.PCF(epi.data, type, hom, het = NULL, r = NULL, correction = "none")
```

### Arguments

epi.data	a three-column numerical matrix that contains coordinates (x and y) for each case and information on case type (e.g. genotype or serotype). First two columns must be x and y
type	an integer giving the column that contains information on case type. Must be an integer or a character
hom	a scalar or vector giving the homotypic case type(s). Equivalent to the 'j' point type used in the cross K function. Must be an integer or character
het	a scalar or vector giving the heterotypic case type(s). Equivalent to the 'i' point type used in the cross K function. The default is NULL, which uses any case type not defined in the hom argument as heterotypic. Must be an integer or a character
r	a numeric vector giving the spatial distances
correction	type of edge correction to be applied (default set to 'none'). See the <a href="#">pcf</a> function in the <b>spatstat.explore</b> package for more details

### Value

a data frame with two columns giving the radius  $r$ , the theoretical value of the Pair Correlation Function for a Poisson process (theo), and value of the Pair Correlation Function pcf

### Author(s)

John Giles

### References

Baddeley A, Rubak E, and Turner R. (2016). "Spatial Point Patterns: Methodology and Applications with R". CRC Press.

**Examples**

```
data(DengueSimR01)

g <- get.cross.PCF(epi.data=DengueSimR01, type=5, hom=2, het=NULL, r=NULL, correction='none')

plot(g$pcf, type='l', xlab='r', ylab='cross PCF')
abline(h=1, col='red', lty=2)
```

get.pi

*Generalized version of get.pi***Description**

Generalized version of the `get.pi` function that takes in an arbitrary function and returns the probability that a point within a particular range of a point of interest shares the relationship specified by the passed in function with that point.

**Usage**

```
get.pi(posmat, fun, r = 1, r.low = rep(0, length(r)), data.frame = TRUE)
```

**Arguments**

posmat	a matrix with columns x, y and any other named columns columns needed by fun
fun	a function that takes in two rows of posmat and returns: <ol style="list-style-type: none"> <li>for pairs included in the numerator and denominator</li> <li>for pairs that should only be included in the denominator</li> <li>for pairs that should be ignored all together</li> </ol> <p>Note that names from posmat are not preserved in calls to fun, so the columns of the matrix should be referenced numerically so this is not available to the fun</p>
r	the series of spatial distances (or there maximums) we are interested in
r.low	the low end of each range, 0 by default
data.frame	logical indicating whether to return results as a data frame (default = TRUE)

**Value**

pi value for each distance range that we look at. Where:

$$\pi(d_1, d_2) = \frac{\sum \mathbf{1}(d_{ij} \in [d_1, d_2]) \mathbf{1}(f(i, j) = 1)}{\sum \sum \mathbf{1}(d_{ij} \in (d_1, d_2)) \mathbf{1}(f(i, j) \in \{1, 2\})}$$

**Author(s)**

Justin Lessler and Henrik Salje

**See Also**

Other get.pi: `get.pi.bootstrap()`, `get.pi.ci()`, `get.pi.permute()`, `get.pi.typed()`, `get.pi.typed.bootstrap()`, `get.pi.typed.permute()`

Other spatialtau: `get.tau()`, `get.theta()`

**Examples**

```
data(DengueSimR02)

r.max<-seq(20,1000,20)
r.min<-seq(0,980,20)

sero.type.func<-function(a,b,tlimit=20){
  if(a[5]==b[5]&(abs(a[3]-b[3])<=tlimit)){rc=1}
  else{rc=2}
  return(rc)
}

sero.pi<-get.pi(DengueSimR02,sero.type.func,r=r.max,r.low=r.min)
```

---

get.pi.bootstrap      *Bootstrap get.pi values.*

---

**Description**

Runs get.pi on multiple bootstraps of the data. Is formulated such that the relationships between points and themselves will not be calculated.

**Usage**

```
get.pi.bootstrap(
  posmat,
  fun,
  r = 1,
  r.low = rep(0, length(r)),
  boot.iter = 500,
  data.frame = TRUE
)
```

**Arguments**

posmat            a matrix with columns type, x and y  
 fun              the function to decide relationships  
 r                 the series of spatial distances we are interested in

r.low	the low end of each range. 0 by default
boot.iter	the number of bootstrap iterations
data.frame	logical indicating whether to return results as a data frame (default = TRUE)

**Value**

pi values for all the distances we looked at

**Note**

In each bootstrap iteration N observations are drawn from the existing data with replacement. To avoid errors in inference resulting from the same observatin being compared with itself in the bootstrapped data set, original indices are perserved, and pairs of points in the bootstrapped dataset with the same original index are ignored.

**Author(s)**

Justin Lessler and Henrik Salje

**See Also**

Other get.pi: [get.pi\(\)](#), [get.pi.ci\(\)](#), [get.pi.permute\(\)](#), [get.pi.typed\(\)](#), [get.pi.typed.bootstrap\(\)](#), [get.pi.typed.permute\(\)](#)

**Examples**

```
#compare normally distributed with uniform points
x<-cbind(1,runif(100,-100,100), runif(100,-100,100))
x<-rbind(x, cbind(2,rnorm(100,0,20), rnorm(100,0,20)))
colnames(x) <- c("type","x","y")

fun<-function(a,b) {
  if(a[1]!=2) return(3)
  if (b[1]==2) return(1)
  return(2)
}

r.max<-seq(10,100,10)
r.min<-seq(0,90,10)
r.mid <- (r.max+r.min)/2

pi<-get.pi(x,fun,r=r.max,r.low=r.min)
pi.boot<-get.pi.bootstrap(x,fun,r=r.max,r.low=r.min,boot.iter=100)

pi.ci<-apply(pi.boot[,-(1:2)],1,quantile,probs=c(0.25,0.75))

plot(r.mid, pi$pi , type="l")
lines(r.mid, pi.ci[1,] , lty=2)
lines(r.mid, pi.ci[2,] , lty=2)
```

---

`get.pi.ci`*Calculate bootstrapped confidence intervals for get.pi values.*

---

### Description

Wrapper to `get.pi.bootstrap` that takes care of calculating the confidence intervals based on the bootstrapped values..

### Usage

```
get.pi.ci(  
  posmat,  
  fun,  
  r = 1,  
  r.low = rep(0, length(r)),  
  boot.iter = 1000,  
  ci.low = 0.025,  
  ci.high = 0.975,  
  data.frame = TRUE  
)
```

### Arguments

<code>posmat</code>	a matrix with columns type, x and y
<code>fun</code>	the function to decide relationships
<code>r</code>	the series of spatial distances wer are interested in
<code>r.low</code>	the low end of each range. 0 by default
<code>boot.iter</code>	the number of bootstrap iterations
<code>ci.low</code>	the low end of the ci...0.025 by default
<code>ci.high</code>	the high end of the ci...0.975 by default
<code>data.frame</code>	logical indicating whether to return results as a data frame (default = TRUE)

### Value

a matrix with a row for the high and low values and a column per distance

### Author(s)

Justin Lessler

### See Also

Other `get.pi`: [get.pi\(\)](#), [get.pi.bootstrap\(\)](#), [get.pi.permute\(\)](#), [get.pi.typed\(\)](#), [get.pi.typed.bootstrap\(\)](#), [get.pi.typed.permute\(\)](#)

**Examples**

```

#compare normally distributed with uniform points
x<-cbind(1,runif(100,-100,100), runif(100,-100,100))
x<-rbind(x, cbind(2,rnorm(100,0,20), rnorm(100,0,20)))
colnames(x) <- c("type","x","y")

fun<-function(a,b) {
  if(a[1]!=2) return(3)
  if (b[1]==2) return(1)
  return(2)
}

r.max<-seq(10,100,10)
r.min<-seq(0,90,10)
r.mid <- (r.max+r.min)/2

pi<-get.pi(x,fun,r=r.max,r.low=r.min)
pi.ci<-get.pi.ci(x,fun,r=r.max,r.low=r.min,boot.iter=100)

plot(r.mid, pi$pi, type="l")
lines(r.mid, pi.ci[,2] , lty=2)
lines(r.mid, pi.ci[,3] , lty=2)

```

---

get.pi.permute

*get the null distribution of the get.pi function*


---

**Description**

Does permutations to calculate the null distribution of get pi if there were no spatial dependence. Randomly reassigs coordinates to each observation permutations times

**Usage**

```

get.pi.permute(
  posmat,
  fun,
  r = 1,
  r.low = rep(0, length(r)),
  permutations,
  data.frame = TRUE
)

```



**Arguments**

posmat	a matrix with columns type, x and y
fun	the function to evaluate
r	the series of spatial distances we are interested in
r.low	the low end of each range....0 by default
permutations	the number of permute iterations
data.frame	logical indicating whether to return results as a data frame (default = TRUE)

**Value**

pi values for all the distances we looked at

**See Also**

Other get.pi: [get.pi\(\)](#), [get.pi.bootstrap\(\)](#), [get.pi.ci\(\)](#), [get.pi.typed\(\)](#), [get.pi.typed.bootstrap\(\)](#), [get.pi.typed.permute\(\)](#)

**Examples**

```
#compare normally distributed with uniform points
x<-cbind(1,runif(100,-100,100), runif(100,-100,100))
x<-rbind(x, cbind(2,rnorm(100,0,20), rnorm(100,0,20)))
colnames(x) <- c("type","x","y")

fun<-function(a,b) {
  if(a[1]!=2) return(3)
  if (b[1]==2) return(1)
  return(2)
}

r.max<-seq(10,100,10)
r.min<-seq(0,90,10)
r.mid <- (r.max+r.min)/2

pi<-get.pi(x, fun, r=r.max, r.low=r.min)
pi.null<-get.pi.permute(x, fun, r=r.max, r.low=r.min, permutations=100)

null.ci<-apply(pi.null[,-(1:2)],1,quantile,probs=c(0.25,0.75))

plot(r.mid, pi$pi, type="l")
lines(r.mid, null.ci[1,] , lty=2)
lines(r.mid, null.ci[2,] , lty=2)
```

---

`get.pi.typed`*Optimized version of get.pi for typed data.*

---

### Description

Version of the `get.pi` function that is optimized for statically typed data. That is data where we are interested in the probability of points within some distance of points of typeA are of typeB.

### Usage

```
get.pi.typed(  
  posmat,  
  typeA = -1,  
  typeB = -1,  
  r = 1,  
  r.low = rep(0, length(r)),  
  data.frame = TRUE  
)
```

### Arguments

<code>posmat</code>	a matrix with columns type, x and y
<code>typeA</code>	the "from" type that we are interested in, -1 is wildcard
<code>typeB</code>	the "to" type that we are interested i, -1 is wildcard
<code>r</code>	the series of spatial distances wer are interested in
<code>r.low</code>	the low end of each range....0 by default
<code>data.frame</code>	logical indicating whether to return results as a data frame (default = TRUE)

### Value

pi values for all the distances we looked at

### Author(s)

Justin Lessler and Henrik Salje

### See Also

Other `get.pi`: [get.pi\(\)](#), [get.pi.bootstrap\(\)](#), [get.pi.ci\(\)](#), [get.pi.permute\(\)](#), [get.pi.typed.bootstrap\(\)](#), [get.pi.typed.permute\(\)](#)

**Examples**

```

data(DengueSimR02)

r.max<-seq(20,1000,20)
r.min<-seq(0,980,20)

#Lets see if there's a difference in spatial dependence by time case occurs
type<-2-(DengueSimR02[,"time"]<120)
tmp<-cbind(DengueSimR02,type=type)

typed.pi<-get.pi.typed(tmp,typeA=1,typeB=2,r=r.max,r.low=r.min)

```

---

```

get.pi.typed.bootstrap
      runs bootstrapping on get.pi.typed

```

---

**Description**

Bootstraps typed pi values. Makes sure distances between a sample and another draw of itself are left out

**Usage**

```

get.pi.typed.bootstrap(
  posmat,
  typeA = -1,
  typeB = -1,
  r = 1,
  r.low = rep(0, length(r)),
  boot.iter,
  data.frame = TRUE
)

```

**Arguments**

posmat	a matrix with columns type, x and y
typeA	the "from" type that we are interested in, -1 is wildcard
typeB	the "to" type that we are interested in, -1 is wildcard
r	the series of spatial distances we are interested in
r.low	the low end of each range....0 by default
boot.iter	the number of bootstrap iterations
data.frame	logical indicating whether to return results as a data frame (default = TRUE)

**Value**

pi values for all the distances we looked at

**See Also**

Other get.pi: `get.pi()`, `get.pi.bootstrap()`, `get.pi.ci()`, `get.pi.permute()`, `get.pi.typed()`, `get.pi.typed.permute()`

**Examples**

```
data(DengueSimR02)

r.max<-seq(20,1000,20)
r.min<-seq(0,980,20)

#Lets see if there's a difference in spatial dependence by time case occurs
type<-2-(DengueSimR02[,"time"]<120)
tmp<-cbind(DengueSimR02,type=type)

typed.pi.bs<-get.pi.typed.bootstrap(tmp,typeA=1,typeB=2,r=r.max,r.low=r.min,boot.iter=100)
```

---

`get.pi.typed.permute` *get the null distribution of the get.pi.typed function*

---

**Description**

Does permutations to calculate the null distribution of get pi if there were no spatial dependence. Randomly reassigns coordinates to each observation permutations times

**Usage**

```
get.pi.typed.permute(
  posmat,
  typeA = -1,
  typeB = -1,
  r = 1,
  r.low = rep(0, length(r)),
  permutations,
  data.frame = TRUE
)
```

**Arguments**

posmat	a matrix with columns type, x and y
typeA	the "from" type that we are interested in, -1 is wildcard
typeB	the "to" type that we are interested in, -1 is wildcard
r	the series of spatial distances we are interested in
r.low	the low end of each range....0 by default
permutations	the number of permute iterations
data.frame	logical indicating whether to return results as a data frame (default = TRUE)

**Value**

pi values for all the distances we looked at

**Author(s)**

Justin Lessler and Henrik Salje

**See Also**

Other get.pi: [get.pi\(\)](#), [get.pi.bootstrap\(\)](#), [get.pi.ci\(\)](#), [get.pi.permute\(\)](#), [get.pi.typed\(\)](#), [get.pi.typed.bootstrap\(\)](#)

**Examples**

```
data(DengueSimR02)

r.max<-seq(20,1000,20)
r.min<-seq(0,980,20)

#Lets see if there's a difference in spatial dependence by time case occurs
type<-2-(DengueSimR02[,"time"]<75)
tmp<-cbind(DengueSimR02,type=type)

typed.pi<-get.pi.typed(tmp,typeA=1,typeB=2,r=r.max,r.low=r.min)
typed.pi.type.null<-get.pi.typed.permute(tmp,typeA=1,typeB=2,r=r.max,r.low=r.min,permutations=100)
```

---

get.tau

*generalized version of get.tau*

---

**Description**

returns the relative probability (or odds) that points at some distance from an index point share some relationship with that point versus the probability (or odds) any point shares that relationship with that point.

**Usage**

```
get.tau(
  posmat,
  fun,
  r = 1,
  r.low = rep(0, length(r)),
  comparison.type = "representative",
  data.frame = TRUE
)
```

**Arguments**

posmat	a matrix with columns x, y and any other named columns columns needed by fun
fun	a function that takes in two rows of posmat and returns: <ol style="list-style-type: none"> <li>1. for pairs included in the numerator (and the denominator for independent data)</li> <li>2. for pairs that should only be included in the denominator</li> <li>3. for pairs that should be ignored all together</li> </ol> <p>Note that names from posmat are not preserved in calls to fun, so the columns of the matrix should be referenced numerically so this is not available to fun</p>
r	the series of spatial distances (or there maximums) we are interested in
r.low	the low end of each range, 0 by default
comparison.type	what type of points are included in the comparison set. <ul style="list-style-type: none"> <li>• "representative" if comparison set is representative of the underlying population</li> <li>• "independent" if comparison set is cases/events coming from an indepent process</li> </ul>
data.frame	logical indicating whether to return results as a data frame (default = TRUE)

**Value**

The tau value for each distance we look at. If comparison.type is "representative", this is:

```
tau = get.pi(posmat, fun, r, r.low)/get.pi(posmat, fun, infinity, 0)
```

If comparison.type is "independent", this is:

```
tau = get.theta(posmat, fun, r, r.low)/get.theta(posmat, fun, infinity, 0)
```

**Author(s)**

Justin Lessler and Henrik Salje

**See Also**

Other get.tau: [get.tau.bootstrap\(\)](#), [get.tau.ci\(\)](#), [get.tau.permute\(\)](#), [get.tau.typed\(\)](#),  
[get.tau.typed.bootstrap\(\)](#), [get.tau.typed.permute\(\)](#)

Other spatialtau: [get.pi\(\)](#), [get.theta\(\)](#)

**Examples**

```

data(DengueSimR01)
data(DengueSimR02)
data(DengueSimRepresentative)

r.max<-seq(20,1000,20)
r.min<-seq(0,980,20)
r.mid<-(r.max+r.min)/2

sero.type.func<-function(a,b,tlimit=20){
  if(a[5]==b[5]&&(abs(a[3]-b[3])<=tlimit)){rc=1}
  else{rc=2}
  return(rc)
}

geno.type.func<-function(a,b,tlimit=20){
  if(a[4]==b[4]&&(abs(a[3]-b[3])<=tlimit)){rc=1}
  else{rc=2}
  return(rc)
}

sero.type.rep.func<-function(a,b,tlimit=20){
  if(a[5]==1&b[5]==1&(abs(a[3]-b[3])<=tlimit)){rc=1}
  else{if(a[5]==1&b[5]==-999){rc=2}else{rc=3}}
  return(rc)
}

sero.tau.R01 <- get.tau(DengueSimR01, sero.type.func, r=r.max, r.low=r.min,
  comparison.type="independent")
geno.tau.R01 <- get.tau(DengueSimR01, geno.type.func, r=r.max, r.low=r.min,
  comparison.type="independent")

sero.tau.R02 <- get.tau(DengueSimR02, sero.type.func, r=r.max, r.low=r.min,
  comparison.type="independent")
geno.tau.R02 <- get.tau(DengueSimR02, geno.type.func, r=r.max, r.low=r.min,
  comparison.type="independent")

sero.tau.representative <- get.tau(DengueSimRepresentative, sero.type.rep.func,
  r=r.max, r.low=r.min, comparison.type="representative")

## R0 of 1
plot(r.mid,sero.tau.R01$tau,ylim=c(0.3,max(geno.tau.R01$tau)),log="y",
  cex.axis=1.25,col=rgb(t(col2rgb("blue")/255),alpha=0.6),
  xlab="Distance (m)",ylab="Tau",cex.main=0.9,lwd=2,type="l",las=1,cex.axis=0.75)
abline(h=1,lty=2)

```

```

abline(v=100,lty=1,lwd=2)
lines(r.mid,geno.tau.R01$tau,pch=20,col=rgb(t(col2rgb("dark green")/255),alpha=0.6),lwd=1)
lines(r.mid,sero.tau.representative$tau,pch=20,col=rgb(t(col2rgb("dark blue")/255),alpha=0.6),lty=2)
legend("topright",
      legend=c("Genotype",
               "Serotype",
               "Serotype (representative population)",
               "Maximum transmission distance"),
      lwd=1,col=c("dark green","blue","blue","black"),
      lty=c(1,1,2,1),bty="n")

## R0 of 2
plot(r.mid,sero.tau.R02$tau,ylim=c(0.3,max(geno.tau.R02)),log="y",
     cex.axis=1.25,col=rgb(t(col2rgb("blue")/255),alpha=0.6),
     xlab="Distance (m)",ylab="Tau",cex.main=0.9,lwd=2,type="l",las=1,cex.axis=0.75)
abline(h=1,lty=2)
abline(v=100,lty=1,lwd=2)
lines(r.mid,geno.tau.R02$tau,pch=20,col=rgb(t(col2rgb("dark green")/255),alpha=0.6),lwd=1)
legend("topright",
      legend=c("Genotype",
               "Serotype",
               "Maximum transmission distance"),
      lwd=1,col=c("dark green","blue","black"),lty=1,bty="n")

```

---

get.tau.bootstrap      *Bootstrap get.tau values.*

---

## Description

Runs get.tau on multiple bootstraps of the data. Is formulated such that the relationship between points and themselves will not be calculated

## Usage

```

get.tau.bootstrap(
  posmat,
  fun,
  r = 1,
  r.low = rep(0, length(r)),
  boot.iter,
  comparison.type = "representative",
  data.frame = TRUE
)

```



**Arguments**

posmat	a matrix appropriate for input to <code>get.tau</code>
fun	a function appropriate as input to <code>get.pi</code>
r	the series of spatial distances we are interested in
r.low	the low end of each range....0 by default
boot.iter	the number of bootstrap iterations
comparison.type	the comparison type to pass as input to <code>get.pi</code>
data.frame	logical indicating whether to return results as a data frame (default = TRUE)

**Value**

a matrix containing all bootstrapped values of tau for each distance interval

**Author(s)**

Justin Lessler and Henrik Salje

**See Also**

Other `get.tau`: [get.tau\(\)](#), [get.tau.ci\(\)](#), [get.tau.permute\(\)](#), [get.tau.typed\(\)](#), [get.tau.typed.bootstrap\(\)](#), [get.tau.typed.permute\(\)](#)

**Examples**

```
#compare normally distributed with uniform points
x<-cbind(1,runif(100,-100,100), runif(100,-100,100))
x<-rbind(x, cbind(2,rnorm(100,0,20), rnorm(100,0,20)))
colnames(x) <- c("type","x","y")

fun<-function(a,b) {
  if(a[1]!=2) return(3)
  if (b[1]==2) return(1)
  return(2)
}

r.max<-seq(10,100,10)
r.min<-seq(0,90,10)
r.mid <- (r.max+r.min)/2

tau<-get.tau(x,fun,r=r.max,r.low=r.min)
tau.boot<-get.tau.bootstrap(x,fun,r=r.max,r.low=r.min,boot.iter=50)

tau.ci<-apply(tau.boot[,-(1:2)],1,quantile,probs=c(0.25,0.75))

plot(r.mid, tau$tau ,ylim=c(min(tau.ci),max(tau.ci)), type="l", log="y")
lines(c(0,100),c(1,1), lty=3, col="grey")
lines(r.mid, tau.ci[1,] , lty=2)
```

```
lines(r.mid, tau.ci[2,] , lty=2)
```

---

```
get.tau.ci
```

```
Bootstrap confidence interval for the get.tau values
```

---

### Description

Wrapper to `get.tau.bootstrap` that takes care of calculating the confidence intervals based on the bootstrapped values

### Usage

```
get.tau.ci(
  posmat,
  fun,
  r = 1,
  r.low = rep(0, length(r)),
  boot.iter = 1000,
  comparison.type = "representative",
  ci.low = 0.025,
  ci.high = 0.975,
  data.frame = TRUE
)
```

### Arguments

<code>posmat</code>	a matrix appropriate for input to <code>get.tau</code>
<code>fun</code>	a function appropriate as input to <code>get.pi</code>
<code>r</code>	the series of spatial distances we are interested in
<code>r.low</code>	the low end of each range...0 by default
<code>boot.iter</code>	the number of bootstrap iterations
<code>comparison.type</code>	the comparison type to pass to <code>get.tau</code>
<code>ci.low</code>	the low end of the ci...0.025 by default
<code>ci.high</code>	the high end of the ci...0.975 by default
<code>data.frame</code>	logical indicating whether to return results as a data frame (default = TRUE)

### Value

a data frame with the point estimate of tau and its low and high confidence interval at each distance

### Author(s)

Justin Lessler and Henrik Salje

**See Also**

Other get.tau: [get.tau\(\)](#), [get.tau.bootstrap\(\)](#), [get.tau.permute\(\)](#), [get.tau.typed\(\)](#), [get.tau.typed.bootstrap\(\)](#), [get.tau.typed.permute\(\)](#)

**Examples**

```
#compare normally distributed with uniform points
x<-cbind(1,runif(100,-100,100), runif(100,-100,100))
x<-rbind(x, cbind(2,rnorm(100,0,20), rnorm(100,0,20)))
colnames(x) <- c("type","x","y")

fun<-function(a,b) {
  if(a[1]!=2) return(3)
  if (b[1]==2) return(1)
  return(2)
}

r.max<-seq(10,100,10)
r.min<-seq(0,90,10)
r.mid <- (r.max+r.min)/2

tau <- get.tau.ci(x,fun,r=r.max,r.low=r.min,boot.iter=50)

plot(r.mid, tau$pt.est, ylim=c(1/max(tau[,3:5]), max(tau[,3:5])), type="l", log="y",
      xlab="Distance", ylab="Tau")
lines(r.mid, tau$ci.low , lty=2)
lines(r.mid, tau$ci.high, lty=2)
lines(c(0,100),c(1,1), lty=3, col="grey")
```

---

get.tau.permute      *get the null distribution of the get.tau function*

---

**Description**

Does permutations to calculate the null distribution of get pi if there were no spatial dependence. Randomly reassigns coordinates to each observation permutations times

**Usage**

```
get.tau.permute(
  posmat,
  fun,
  r = 1,
  r.low = rep(0, length(r)),
  permutations,
  comparison.type = "representative",
```

```

    data.frame = TRUE
  )

```

### Arguments

posmat	a matrix appropriate for input to <code>get.tau</code>
fun	a function appropriate for input to <code>get.tau</code>
r	the series of spatial distances we are interested in
r.low	the low end of each range....0 by default
permutations	the number of permute iterations
comparison.type	the comparison type to pass as input to <code>get.pi</code>
data.frame	logical indicating whether to return results as a data frame (default = TRUE)

### Value

tau values for all the distances we looked at

### Author(s)

Justin Lessler and Henrik Salje

### See Also

Other `get.tau`: [get.tau\(\)](#), [get.tau.bootstrap\(\)](#), [get.tau.ci\(\)](#), [get.tau.typed\(\)](#), [get.tau.typed.bootstrap\(\)](#), [get.tau.typed.permute\(\)](#)

### Examples

```

#compare normally distributed with uniform points
x<-cbind(1,runif(100,-100,100), runif(100,-100,100))
x<-rbind(x, cbind(2,rnorm(100,0,20), rnorm(100,0,20)))
colnames(x) <- c("type","x","y")

fun<-function(a,b) {
  if(a[1]!=2) return(3)
  if (b[1]==2) return(1)
  return(2)
}

r.max<-seq(10,100,10)
r.min<-seq(0,90,10)
r.mid <- (r.max+r.min)/2

tau<-get.tau(x,fun,r=r.max,r.low=r.min,comparison.type = "independent")
tau.null<-get.tau.permute(x,fun,r=r.max,r.low=r.min,permutations=50,comparison.type = "independent")

null.ci<-apply(tau.null[,-(1:2)],1,quantile,probs=c(0.25,0.75))

```

```

plot(r.mid, tau$tau, ylim=c(1/max(tau$tau),max(tau$tau)), type="l", log="y")
lines(c(0,100),c(1,1), lty=3, col="grey")
lines(r.mid, null.ci[1,] , lty=2)
lines(r.mid, null.ci[2,] , lty=2)

```

---

get.tau.typed

*Optimized version of get.tau for typed data*


---

### Description

Version of the get.tau function that is optimized for statically typed data. That is data where we want the relationship between points of type A and points of type B

### Usage

```

get.tau.typed(
  posmat,
  typeA = -1,
  typeB = -1,
  r = 1,
  r.low = rep(0, length(r)),
  comparison.type = "representative",
  data.frame = TRUE
)

```

### Arguments

posmat	a matrix with columns type, x and y
typeA	the "from" type that we are interested in, -1 is wildcard
typeB	the "to" type that we are interested in, -1 is wildcard
r	the series of spatial distances we are interested in
r.low	the low end of each range....0 by default
comparison.type	what type of points are included in the comparison set. <ul style="list-style-type: none"> <li>"representative" if comparison set is representative of the underlying population</li> <li>"independent" if comparison set is cases/events coming from an independent process</li> </ul>
data.frame	logical indicating whether to return results as a data frame (default = TRUE)

### Value

data frame of tau values for all the distances

**Author(s)**

Justin Lessler and Henrik Salje

**See Also**

Other get.tau: [get.tau\(\)](#), [get.tau.bootstrap\(\)](#), [get.tau.ci\(\)](#), [get.tau.permute\(\)](#), [get.tau.typed.bootstrap\(\)](#), [get.tau.typed.permute\(\)](#)

**Examples**

```
data(DengueSimR02)

r.max<-seq(20,1000,20)
r.min<-seq(0,980,20)
r.mid<-(r.max+r.min)/2

#Lets see if there's a difference in spatial dependence by time case occurs
type<-2-(DengueSimR02[,"time"]<120)
tmp<-cbind(DengueSimR02,type=type)

typed.tau<-get.tau.typed(tmp,typeA=1,typeB=2,r=r.max,r.low=r.min,comparison.type = "independent")

plot(r.mid,typed.tau$tau,log="y",cex.axis=1.25,
      xlab="Distance (m)",ylab="Tau",cex.main=0.9,lwd=2,type="l")
abline(h=1,lty=2)
```

---

```
get.tau.typed.bootstrap
      runs bootstrapping for get.tau.typed
```

---

**Description**

runs bootstrapping for get.tau.typed

**Usage**

```
get.tau.typed.bootstrap(
  posmat,
  typeA = -1,
  typeB = -1,
  r = 1,
  r.low = rep(0, length(r)),
  boot.iter,
  comparison.type = "representative",
  data.frame = TRUE
)
```

**Arguments**

posmat	a matrix with columns type, x and y
typeA	the "from" type that we are interested in, -1 is wildcard
typeB	the "to" type that we are interested i, -1 is wildcard
r	the series of spatial distances we are interested in
r.low	the low end of each range....0 by default
boot.iter	the number of bootstrap iterations
comparison.type	what type of points are included in the comparison set. <ul style="list-style-type: none"> <li>• "representative" if comparison set is representative of the underlying population</li> <li>• "independent" if comparison set is cases/events coming from an independent process</li> </ul>
data.frame	logical indicating whether to return results as a data frame (default = TRUE)

**Value**

tau values for all the distances we looked at

**Author(s)**

Justin Lessler and Henrik Salje

**See Also**

Other get.tau: [get.tau\(\)](#), [get.tau.bootstrap\(\)](#), [get.tau.ci\(\)](#), [get.tau.permute\(\)](#), [get.tau.typed\(\)](#), [get.tau.typed.permute\(\)](#)

**Examples**

```
data(DengueSimulationR02)

r.max<-seq(20,1000,20)
r.min<-seq(0,980,20)
r.mid<-(r.max+r.min)/2

# Lets see if there's a difference in spatial dependence between those that occurred
# late versus early in the outbreak

type <- 2 - (DengueSimR02[,"time"] < 120)
tmp <- cbind(DengueSimR02, type=type)

typed.tau <- get.tau.typed(tmp, typeA=1, typeB=2, r=r.max, r.low=r.min,
                          comparison.type = "independent")

typed.tau.type.bs <- get.tau.typed.bootstrap(tmp, typeA=1, typeB=2, r=r.max, r.low=r.min,
                                             boot.iter=100, comparison.type = "independent")
```

```

ci <- apply(typed.tau.type.bs[,-(1:2)], 1, quantile, probs=c(0.025,0.975))

plot(r.mid, typed.tau$tau, log="y",
     ylim=c(0.1,4), cex.axis=1.25,
     xlab="Distance (m)", ylab="Tau",
     cex.main=0.9, lwd=2, type="n")
abline(h=1,lty=1)
lines(r.mid,typed.tau$tau,pch=20,col=1,lwd=3)
lines(r.mid, ci[1,] , lty=2)
lines(r.mid, ci[2,] , lty=2)

```

---

get.tau.typed.permute *get the null distribution for the get.tau.typed function*

---

## Description

get the null distribution for the get.tau.typed function

## Usage

```

get.tau.typed.permute(
  posmat,
  typeA = -1,
  typeB = -1,
  r = 1,
  r.low = rep(0, length(r)),
  permutations,
  comparison.type = "representative",
  data.frame = TRUE
)

```

## Arguments

posmat	a matrix with columns type, x and y
typeA	the "from" type that we are interested in, -1 is wildcard
typeB	the "to" type that we are interested i, -1 is wildcard
r	the series of spatial distances we are interested in
r.low	the low end of each range....0 by default
permutations	the number of permute iterations
comparison.type	what type of points are included in the comparison set. <ul style="list-style-type: none"> <li>"representative" if comparison set is representative of the underlying population</li> </ul>



- "independent" if comparison set is cases/events coming from an independent process

data.frame      logical indicating whether to return results as a data frame (default = TRUE)

### Value

a matrix with permutation tau values for each distance specified

### Author(s)

Justin Lessler and Henrik Salje

### See Also

Other get.tau: [get.tau\(\)](#), [get.tau.bootstrap\(\)](#), [get.tau.ci\(\)](#), [get.tau.permute\(\)](#), [get.tau.typed\(\)](#), [get.tau.typed.bootstrap\(\)](#)

### Examples

```
data(DengueSimulationR02)

r.max<-seq(20,1000,20)
r.min<-seq(0,980,20)
r.mid<-(r.max+r.min)/2

#Lets see if there's a difference in spatial dependence by time case occurs
type <- 2 - (DengueSimR02[,"time"] < 120)
tmp <- cbind(DengueSimR02, type=type)

typed.tau <- get.tau.typed(tmp, typeA=1, typeB=2, r=r.max, r.low=r.min,
                           comparison.type = "independent")

typed.tau.type.null<-get.tau.typed.permute(tmp, typeA=1, typeB=2, r=r.max, r.low=r.min,
                                           permutations=100, comparison.type = "independent")

null.ci <- apply(typed.tau.type.null[,-(1:2)], 1, quantile, probs=c(0.025,0.975))

plot(r.mid, typed.tau$tau, ylim=c(0.3,4), log="y", cex.axis=1.25,
      xlab="Distance (m)", ylab="Tau", cex.main=0.9, lwd=2, type="n")
abline(h=1,lty=1)
lines(r.mid,typed.tau$tau,pch=20,col=1,lwd=3)
lines(r.mid, null.ci[1,] , lty=2)
lines(r.mid, null.ci[2,] , lty=2)
```

---

<code>get.theta</code>	<i>Generalized version of <code>get.theta</code></i>
------------------------	--

---

### Description

Generalized version of the `get.theta` function that takes in an arbitrary function and returns the odds that a point within a particular range of a point of interest shares the relationship specified by the passed in function with that point.

### Usage

```
get.theta(posmat, fun, r = 1, r.low = rep(0, length(r)), data.frame = TRUE)
```

### Arguments

<code>posmat</code>	a matrix with columns x, y and any other named columns columns needed by fun
<code>fun</code>	a function that takes in two rows of <code>posmat</code> and returns: <ol style="list-style-type: none"> <li>1. for pairs that are (potentially) related</li> <li>2. for pairs that are unrelated</li> <li>3. for pairs that should be ignored all together</li> </ol> <p>Note that names from <code>posmat</code> are not preserved in calls to <code>fun</code>, so the columns of the matrix should be referenced numerically so this is not available to the fun</p>
<code>r</code>	the series of spatial distances (or there maximums) we are interested in
<code>r.low</code>	the low end of each range, 0 by default
<code>data.frame</code>	logical indicating whether to return results as a data frame (default = TRUE)

### Value

theta value for each distance range that we look at. Where:

$$\theta(d_1, d_2) = \frac{\sum \mathbf{1}_{d_{ij} \in [d_1, d_2)} \mathbf{1}(f(i, j) = 1)}{\sum \sum \mathbf{1}_{d_{ij} \in [d_1, d_2)} \mathbf{1}(f(i, j) = 2)}$$

### Author(s)

Justin Lessler and Henrik Salje

### See Also

Other `get.theta`: [get.theta.bootstrap\(\)](#), [get.theta.ci\(\)](#), [get.theta.permute\(\)](#), [get.theta.typed\(\)](#), [get.theta.typed.bootstrap\(\)](#), [get.theta.typed.permute\(\)](#)

Other `spatialtau`: [get.pi\(\)](#), [get.tau\(\)](#)

**Examples**

```

data(DengueSimR02)

r.max<-seq(20,1000,20)
r.min<-seq(0,980,20)

sero.type.func<-function(a,b,tlimit=20){
  if(a[5]==b[5]&(abs(a[3]-b[3])<=tlimit){rc=1}
  else{rc=2}
  return(rc)
}

sero.theta<-get.theta(DengueSimR02,sero.type.func,r=r.max,r.low=r.min)

```

---

get.theta.bootstrap    *Bootstrap get.theta values.*

---

**Description**

Runs get.theta on multiple bootstraps of the data. Is formulated such that the relationships between points and themselves will not be calculated.

**Usage**

```

get.theta.bootstrap(
  posmat,
  fun,
  r = 1,
  r.low = rep(0, length(r)),
  boot.iter = 500,
  data.frame = TRUE
)

```

**Arguments**

posmat	a matrix with columns type, x and y
fun	the function to decide relationships
r	the series of spatial distances we are interested in
r.low	the low end of each range. 0 by default
boot.iter	the number of bootstrap iterations
data.frame	logical indicating whether to return results as a data frame (default = TRUE)

**Value**

theta values for all the distances we looked at

**Note**

In each bootstrap iteration N observations are drawn from the existing data with replacement. To avoid errors in inference resulting from the same observatin being compared with itself in the bootstrapped data set, original indices are perserved, and pairs of points in the bootstrapped dataset with the same original index are ignored.

**Author(s)**

Justin Lessler and Henrik Salje

**See Also**

Other `get.theta`: [get.theta\(\)](#), [get.theta.ci\(\)](#), [get.theta.permute\(\)](#), [get.theta.typed\(\)](#), [get.theta.typed.bootstrap\(\)](#), [get.theta.typed.permute\(\)](#)

**Examples**

```
#compare normally distributed with uniform points
x<-cbind(1,runif(100,-100,100), runif(100,-100,100))
x<-rbind(x, cbind(2,rnorm(100,0,20), rnorm(100,0,20)))
colnames(x) <- c("type","x","y")

fun<-function(a,b) {
  if(a[1]!=2) return(3)
  if (b[1]==2) return(1)
  return(2)
}

r.max<-seq(10,100,10)
r.min<-seq(0,90,10)
r.mid <- (r.max+r.min)/2

theta<-get.theta(x, fun, r=r.max, r.low=r.min)
theta.boot<-get.theta.bootstrap(x, fun, r=r.max, r.low=r.min, boot.iter=100)

theta.ci<-apply(theta.boot[, -(1:2)], 1, quantile, probs=c(0.25, 0.75))

plot(r.mid, theta$theta , type="l")
lines(r.mid, theta.ci[1,] , lty=2)
lines(r.mid, theta.ci[2,] , lty=2)
```

---

get.theta.ci	Calculate bootstrapped confidence intervals for get.theta values.
--------------	---

---

### Description

Wrapper to `get.theta.bootstrap` that takes care of calculating the confidence intervals based on the bootstrapped values.

### Usage

```
get.theta.ci(  
  posmat,  
  fun,  
  r = 1,  
  r.low = rep(0, length(r)),  
  boot.iter = 1000,  
  ci.low = 0.025,  
  ci.high = 0.975,  
  data.frame = TRUE  
)
```

### Arguments

<code>posmat</code>	a matrix with columns type, x and y
<code>fun</code>	the function to decide relationships
<code>r</code>	the series of spatial distances we are interested in
<code>r.low</code>	the low end of each range. 0 by default
<code>boot.iter</code>	the number of bootstrap iterations
<code>ci.low</code>	the low end of the ci...0.025 by default
<code>ci.high</code>	the high end of the ci...0.975 by default
<code>data.frame</code>	logical indicating whether to return results as a data frame (default = TRUE)

### Value

a matrix with a row for the high and low values and a column per distance

### Author(s)

Justin Lessler

### See Also

Other `get.theta`: [get.theta\(\)](#), [get.theta.bootstrap\(\)](#), [get.theta.permute\(\)](#), [get.theta.typed\(\)](#), [get.theta.typed.bootstrap\(\)](#), [get.theta.typed.permute\(\)](#)

**Examples**

```
#compare normally distributed with uniform points
x<-cbind(1,runif(100,-100,100), runif(100,-100,100))
x<-rbind(x, cbind(2,rnorm(100,0,20), rnorm(100,0,20)))
colnames(x) <- c("type","x","y")

fun<-function(a,b) {
  if(a[1]!=2) return(3)
  if (b[1]==2) return(1)
  return(2)
}

r.max<-seq(10,100,10)
r.min<-seq(0,90,10)
r.mid <- (r.max+r.min)/2

theta<-get.theta(x, fun,r=r.max,r.low=r.min)
theta.ci<-get.theta.ci(x, fun,r=r.max,r.low=r.min,boot.iter=100)

plot(r.mid, theta$theta, type="l")
lines(r.mid, theta.ci[,2] , lty=2)
lines(r.mid, theta.ci[,3] , lty=2)
```

---

get.theta.permute      *get the null distribution of the get.theta function*

---

**Description**

Does permutations to calculate the null distribution of get theta if there were no spatial dependence. Randomly reassigned coordinates to each observation permutations times

**Usage**

```
get.theta.permute(
  posmat,
  fun,
  r = 1,
  r.low = rep(0, length(r)),
  permutations,
  data.frame = TRUE
)
```

**Arguments**

posmat                  a matrix with columns type, x and y

fun	the function to evaluate
r	the series of spatial distances we are interested in
r.low	the low end of each range....0 by default
permutations	the number of permute iterations
data.frame	logical indicating whether to return results as a data frame (default = TRUE)

**Value**

theta values for all the distances we looked at

**See Also**

Other get.theta: [get.theta\(\)](#), [get.theta.bootstrap\(\)](#), [get.theta.ci\(\)](#), [get.theta.typed\(\)](#), [get.theta.typed.bootstrap\(\)](#), [get.theta.typed.permute\(\)](#)

**Examples**

```
#compare normally distributed with uniform points
x<-cbind(1,runif(100,-100,100), runif(100,-100,100))
x<-rbind(x, cbind(2,rnorm(100,0,20), rnorm(100,0,20)))
colnames(x) <- c("type","x","y")

fun<-function(a,b) {
  if(a[1]!=2) return(3)
  if (b[1]==2) return(1)
  return(2)
}

r.max<-seq(10,100,10)
r.min<-seq(0,90,10)
r.mid <- (r.max+r.min)/2

theta<-get.theta(x,fun,r=r.max,r.low=r.min)
theta.null<-get.theta.permute(x,fun,r=r.max,r.low=r.min,permutations=100)

null.ci<-apply(theta.null[,-(1:2)],1,quantile,probs=c(0.25,0.75))

plot(r.mid, theta$theta , type="l")
lines(r.mid, null.ci[1,] , lty=2)
lines(r.mid, null.ci[2,] , lty=2)
```

---

get.theta.typed      *Optimized version of get.theta for typed data.*

---

### Description

Version of the get.theta function that is optimized for statically typed data. That is data where we are interested in the odds that points within some distance of points of typeA are of typeB.

### Usage

```
get.theta.typed(  
  posmat,  
  typeA = -1,  
  typeB = -1,  
  r = 1,  
  r.low = rep(0, length(r)),  
  data.frame = TRUE  
)
```

### Arguments

posmat	a matrix with columns type, x and y
typeA	the "from" type that we are interested in, -1 is wildcard
typeB	the "to" type that we are interested i, -1 is wildcard
r	the series of spatial distances wer are interested in
r.low	the low end of each range....0 by default
data.frame	logical indicating whether to return results as a data frame (default = TRUE)

### Value

theta values for all the distances we looked at

### Author(s)

Justin Lessler and Henrik Salje

### See Also

Other get.theta: [get.theta\(\)](#), [get.theta.bootstrap\(\)](#), [get.theta.ci\(\)](#), [get.theta.permute\(\)](#), [get.theta.typed.bootstrap\(\)](#), [get.theta.typed.permute\(\)](#)



**Examples**

```

data(DengueSimR02)

r.max<-seq(20,1000,20)
r.min<-seq(0,980,20)

#Lets see if there's a difference in spatial dependence by time case occurs
type<-2-(DengueSimR02[,"time"]<120)
tmp<-cbind(DengueSimR02,type=type)

typed.theta.R01<-get.theta.typed(tmp,typeA=2,typeB=2,r=r.max,r.low=r.min)

```

---

```

get.theta.typed.bootstrap
      runs bootstrapping on get.theta.typed

```

---

**Description**

Bootstraps typed pi values. Makes sure distances between a sample and another draw of itself are left out

**Usage**

```

get.theta.typed.bootstrap(
  posmat,
  typeA = -1,
  typeB = -1,
  r = 1,
  r.low = rep(0, length(r)),
  boot.iter,
  data.frame = TRUE
)

```

**Arguments**

posmat	a matrix with columns type, x and y
typeA	the "from" type that we are interested in, -1 is wildcard
typeB	the "to" type that we are interested in, -1 is wildcard
r	the series of spatial distances we are interested in
r.low	the low end of each range....0 by default
boot.iter	the number of bootstrap iterations
data.frame	logical indicating whether to return results as a data frame (default = TRUE)

**Value**

theta values for all the distances we looked at

**See Also**

Other `get.theta`: [get.theta\(\)](#), [get.theta.bootstrap\(\)](#), [get.theta.ci\(\)](#), [get.theta.permute\(\)](#), [get.theta.typed\(\)](#), [get.theta.typed.permute\(\)](#)

**Examples**

```
data(DengueSimR01)

r.max<-seq(20,1000,20)
r.min<-seq(0,980,20)

#Lets see if there's a difference in spatial dependence by time case occurs
type<-2-(DengueSimR01[,"time"]<75)
tmp<-cbind(DengueSimR01,type=type)

typed.theta.bs<-get.theta.typed.bootstrap(tmp,typeA=1,typeB=2,r=r.max,r.low=r.min,boot.iter=100)
```

---

```
get.theta.typed.permute
```

*get the null distribution of the `get.theta.typed` function*

---

**Description**

Does permutations to calculate the null distribution of `get.theta` if there were no spatial dependence. Randomly reassigns coordinates to each observation permutations times

**Usage**

```
get.theta.typed.permute(
  posmat,
  typeA = -1,
  typeB = -1,
  r = 1,
  r.low = rep(0, length(r)),
  permutations,
  data.frame = TRUE
)
```

**Arguments**

<code>posmat</code>	a matrix with columns type, x and y
<code>typeA</code>	the "from" type that we are interested in, -1 is wildcard
<code>typeB</code>	the "to" type that we are interested i, -1 is wildcard
<code>r</code>	the series of spatial distances we are interested in
<code>r.low</code>	the low end of each range....0 by default
<code>permutations</code>	the number of permute iterations
<code>data.frame</code>	logical indicating whether to return results as a data frame (default = TRUE)

**Value**

theta values for all the distances we looked at

**Author(s)**

Justin Lessler and Henrik Salje

**See Also**

Other `get.theta`: [get.theta\(\)](#), [get.theta.bootstrap\(\)](#), [get.theta.ci\(\)](#), [get.theta.permute\(\)](#), [get.theta.typed\(\)](#), [get.theta.typed.bootstrap\(\)](#)

**Examples**

```
data(DengueSimR01)

r.max<-seq(20,1000,20)
r.min<-seq(0,980,20)

#Lets see if there's a difference in spatial dependence by time case occurs
type<-2-(DengueSimR01[,"time"]<75)
tmp<-cbind(DengueSimR01,type=type)

typed.theta.R01<-get.theta.typed(tmp,typeA=1,typeB=2,r=r.max,r.low=r.min)
typed.theta.type.null<-get.theta.typed.permute(tmp, typeA=1, typeB=2,
                                              r=r.max, r.low=r.min, permutations=100)
```

---

`get.transdist.theta`    *Get weights of transmission distance theta*

---

### Description

This function estimates the weights of each theta (number of transmission events separating cases at two time points). A randomized transmission tree is drawn and the number of transmission events separating cases at two time points is calculated based on probabilities found in the Wallinga-Teunis matrix.

### Usage

```
get.transdist.theta(  
  wal.teun.mat,  
  cases,  
  gen.t.mean,  
  max.sep,  
  ret.theta.mat = FALSE  
)
```

### Arguments

<code>wal.teun.mat</code>	a Wallinga-Teunis matrix produced by the <code>est.wt.matrix</code> function
<code>cases</code>	a vector of case times for each case
<code>gen.t.mean</code>	the mean generation time of the infecting pathogen
<code>max.sep</code>	maximum number of transmission events allowed between two cases
<code>ret.theta.mat</code>	logical value which returns the matrix of estimated theta values (default = FALSE)

### Value

a three-dimensional array containing normalized theta weights. Columns and rows represent unique case times. The third dimension is the number of transmission events between two cases.

### Author(s)

John Giles, Justin Lessler, and Henrik Salje

### References

Salje H, Cummings DAT and Lessler J (2016). “Estimating infectious disease transmission distances using the overall distribution of cases.” *Epidemics*, 17, pp. 10–18. ISSN 1755-4365, doi: [10.1016/j.epidem.2016.10.001](https://doi.org/10.1016/j.epidem.2016.10.001).

### See Also

Other transdist: [est.transdist\(\)](#), [est.transdist.bootstrap.ci\(\)](#), [est.transdist.temporal\(\)](#), [est.transdist.temporal.bootstrap.ci\(\)](#), [est.transdist.theta.weights\(\)](#)

**Examples**

```

case.times <- c(1,2,2,3,3)
gen <- c(0, 2/3, 1/3, 0, 0)
t.density <- gen/sum(gen)

gen.time <- 2 # mean generation time

wt <- est.wt.matrix(case.times=case.times, gen.t.dist=t.density)

ngen <- round((max(case.times) - min(case.times)) / gen.time) + 1 # Number of generations

a <- get.transdist.theta(wal.teun.mat=wt,
                        cases=case.times,
                        gen.t.mean=gen.time,
                        max.sep=ngen*2)

```

---

sim.epidemic

*Simulation of an epidemic in space and time*


---

**Description**

A function which simulates the spatial spread of infections through time given the reproductive number ( $R$ ), a function describing the spatial transmission kernel (`trans.kern.func`), and the mean and standard deviation of the generation time distribution (`gen.t.mean` and `gen.t.sd`) for the infecting pathogen. The function returns the location ( $x$ ,  $y$ ) and time ( $t$ ) for each case of infection in the simulation.

**Usage**

```

sim.epidemic(
  R,
  gen.t.mean,
  gen.t.sd,
  trans.kern.func,
  tot.generations = 10,
  min.cases = 0,
  max.try = 1000
)

```

**Arguments**

<code>R</code>	a scalar or a vector of length <code>tot.generations</code> providing the reproductive number for the epidemic. If scalar, the $R$ value is constant. If a vector, the $R$ value varies according to each generation in the vector.
<code>gen.t.mean</code>	mean of generation time

`gen.t.sd`            standard deviation of the generation time (assumed to be normally distributed)  
`trans.kern.func`       a function for the transmission kernel that takes `n` as an argument. Function and associated parameters must be given in a list object.  
`tot.generations`       the total number of generations in the epidemic, where the index case (`x,y,t = [0,0,0]`) is considered generation zero (default = 10)  
`min.cases`            the minimum number of cases in the epidemic (default = 0)  
`max.try`              maximum number of tries to achieve the minimum number of cases (default = 1000)

**Value**

a numerical matrix with three columns giving the coordinates `x` and `y`, and time `t` of simulated cases

**Author(s)**

John Giles, Justin Lessler, and Henrik Salje

**Examples**

```

set.seed(1)

dist.func <- alist(n=1, a=1/100, rexp(n, a)) # Exponential transmission kernel with mean = sd = 100

# Simulate epidemic with constant R value
a <- sim.epidemic(R=1.5,
  gen.t.mean=7,
  gen.t.sd=2,
  tot.generations=15,
  min.cases=100,
  trans.kern.func=dist.func)

sim.plot(a)

# Simulate an epidemic with variable R value
r1 <- 2
r2 <- 0.25
tg <- 25
R <- seq(r1, r2, (r2 - r1)/(tg - 1))

b <- sim.epidemic(R=R,
  gen.t.mean=7,
  gen.t.sd=2,
  tot.generations=tg,
  min.cases=100,
  trans.kern.func=dist.func)

sim.plot(b)

```

---

<code>sim.plot</code>	<i>Plot output of simulated epidemic</i>
-----------------------	--

---

**Description**

A simple visualization function which plots the location of the index case and the spatial distribution of subsequent cases, and the epidemic curve showing the case count over time.

**Usage**

```
sim.plot(sim)
```

**Arguments**

`sim` a three-column matrix object produced by the `sim.epidemic` function

**Value**

A two-panel plotted object

**Author(s)**

John Giles, Justin Lessler, and Henrik Salje

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